

## Protocol Calbindin D28 - ELISA

### **Products**

30% H<sub>2</sub>O<sub>2</sub> solution: Merck, 8222887.1000  
3,3',5,5'-tetramethylbenzidin (TMB): Fluka, 87748  
Aceton: Fluka, 00570  
Bovine serum albumin: Fluka, 05480  
Citric acid monohydrate: Fluka, 27490  
Ethanol: Fluka, 02860  
Kathon MW/WT: Christ AG, Aesch, Switzerland  
NaHCO<sub>3</sub>: Fluka, 71628  
Phenol: Merck, 206  
Sulfuric acid 97%: Fluka, 84720  
Tris/HCl (Tris-(hydroxymethyl)-aminoethan): Fluka, 93352  
Tween-20: Fluka, 93773

### **Reagents**

#### **REAGENT 1. Rabbit CB 38 antibody solution**

- Tris/HCl pH 7.5 20 mmol/L
- NaCl 150 mmol/L
- Kathon MW/WT 0.2 ml/L
- Amaranth 10 mg/L
- **Added only before use:** rabbit CB 38 antibody (1:5 - 1:20)

*This antibody solution is used for sensitization of micro-well plate. Preparation: CB 38 antibody (globulin fraction) is added ONLY directly before running the test at a concentration of 1:5 - 1:20! REAGENT 1 will be diluted at 1:1'000 in SOLUTION #3.1. -> end concentration of the antibody 1:5'000 - 1:20'000. Stability: Kathon allows this solution to be stored at least 6 month at 2-8°C without significant loss of immunological activity*

#### **REAGENT 2. Mouse CB 300 antibody solution**

- Tris/HCl pH 7.5 0.1 mol/L
- Bovine serum albumin (BSA) 5.0 g/L
- Phenol 1.0 g/L
- Kathon (MW/WT) 0.2 ml/L
- Evans Blue 10 mg/L
- **Added only before use:** mouse CB 300 antibody (1:10 - 1:20)

*Preparation: monoclonal mouse CB 300 antibody is added ONLY directly before running the test at a concentration of 1:10 - 1:20! REAGENT 2 will be diluted 1:500 in REAGENT 4 -> end concentration of the antibody 1:5'000 - 1:10'000. Stability: at least 6 months at 2-8°C*

#### **REAGENT 3. Calbindin D28 standard solution**



1000 ng/mL recombinant rat Calbindin D28 in:

- Potassium-phosphate pH 6.5 0.1 mol/L
- Bovine serum albumin (BSA) 5.0 g/L
- Kathon MW/WT 0.2 ml/L

*The recombinant Calbindin D28 is ONLY added to the solution directly before use at a concentration of 1ug/ml!*

*Stability: at least 6 months at 2-8°C*

#### **REAGENT 4. Test dilution buffer**

- Tris/HCl pH 7.5 0.2 mol/L
- Bovine serum albumin (BSA) 5.0 g/L
- Kathon MW/WT 0.2 ml/L
- Phenol 1.0 g/L

*Preparation: add 6.05 g of Tris/HCl, 0.1 ml kathon MW/WT, 0.5 g phenol and 2.5 BSA in 400 ml ddH<sub>2</sub>O. Adjust pH to 7.5 with 4 N HCl, and complete the volume to 500 ml with ddH<sub>2</sub>O.*

*Stability: at least 6 months at 2-8°C*

#### **REAGENT 5. Tetramethylbenzidin-H<sub>2</sub>O<sub>2</sub> solution (TMB)**

- 3,3',5,5'-tetramethylbenzidin 20 mmol/L
- Aceton 100 ml/L
- Ethanol 900 ml/L
- H<sub>2</sub>O<sub>2</sub> 50 mmol/L

*Preparation: dissolve 240 mg TMB in 5 ml aceton. Further add 45 ml ethanol and then 300 µl of 30% H<sub>2</sub>O<sub>2</sub> solution.*

*Stability: at least 6 month at 15-25°, in a well closed, light-protected glass container*

#### **REAGENT 6. Substrate buffer**

- Potassium-citrate buffer pH 4.1 30 mmol/L
- Kathon MW/WT 0.2 ml/L

*Preparation: dissolve 6.3 g citric acid monohydrate in about 800 ml ddH<sub>2</sub>O.*

*Adjust pH to 4.1 with 4 N KOH. Complete the volume to 1000 ml with ddH<sub>2</sub>O and add 200 µl kathon MW/WT*

*Stability: Few months at 15-25°C*

#### **Other solutions**

##### **Sensitization buffer (SOLUTION #3.1.)**

- NaHCO<sub>3</sub> pH 9.5 100 mmol/L

*Preparation: dissolve 4.2 g NaHCO<sub>3</sub> in 400 ml ddH<sub>2</sub>O. Adjust pH to 9.5 with 4 N NaOH. Complete the volume to 500 ml with ddH<sub>2</sub>O*

*Stability: at least 6 months at 2-8°C*

##### **Blocking buffer (SOLUTION #3.2.)**



- Tris/HCl pH 7.5 200 mmol/L
- Bovine serum albumin (BSA) 10 g/L
- Kathon MW/WT 0.2 ml/L

*Preparation: dissolve 12.1 g of Tris-HCl and 0.1 ml Kathon MW/WT in 400 ml ddH<sub>2</sub>O. Adjust pH to 7.5 with 4 N HCl. Further add 5 g BSA. Complete the volume to 500 ml with ddH<sub>2</sub>O*

*Stability: at least 6 months at 2-8°C*

### **Stop buffer (SOLUTION #3.3.)**

- Sulfuric acid 1.0 mol/L

*Preparation: add 28 ml sulfuric acid to 500 ml ddH<sub>2</sub>O*

→ *Warning: do not add water in acid*

### **Washing buffer (SOLUTION #3.4.)**

- Tween-20 0.5 ml/L

*Preparation: dilute 500 µl Tween-20 in 1000 ml ddH<sub>2</sub>O*

### **Other materials**

**Micro-well plates:** polystyrol, flat bottom, 96-wells, NUNC-immunoplate, MAXISORP, 4-42 404

**Multichannel photometer:** filter 450 nm

### **ELISA running**

#### **Sensitization of micro-well plate with rabbit CB 38 antibody**

20 ml sensitization solution is required per micro-well plate

- A corresponding amount of antibody solution (**REAGENT 1**) is diluted 1:1000 in sensitization buffer (**SOLUTION #3.1.**): for example 20 µl **REAGENT 1** in 20 ml **SOLUTION #3.1.**  
→ rabbit CB 38 antibody (end concentration 1:5'000 - 1:20'000) in 100 mmol/L NaHCO<sub>3</sub> pH 9.5
- Pipette 200 µl of this solution per well
- Incubate 16-24 hrs at 15-25°C
- Remove sensitization solution and wash 3x with ddH<sub>2</sub>O
- Pipette 200 µl of blocking buffer (**SOLUTION #3.2.**) per well
- Close the plate with parafilm and incubate at least one day at 15-25°C

The micro-well plates sensitized with CB 38 antibody can be stored (under humidified conditions at 2-8°C) in the blocking buffer until 2 months without significant loss of immunological binding capacity

#### **Preparation of solutions for standard curve and loading of samples**

- Dilute Calbindin D28 standard solution 1000 ng/ml (**REAGENT 3**) in test dilution buffer (**REAGENT 4**)  
→ 1:200 for 5.0 ng/ml  
→ 1:400 for 2.5 ng/L  
→ 1:800 for 1.25 ng/L

→ The null-value is given by the test dilution buffer alone

- h) Prepare mouse CB 300 antibody test solution: Dilute mouse CB 300 antibody solution (**REAGENT 2**) 1:500 in test dilution buffer (**REAGENT 4**) (mouse CB 300 antibody end concentration 1:5'000 - 1:10'000)

5 ml of this preparation is required per micro-well plate (5 ml **REAGENT 4** and 5 µl **REAGENT 2**)

- i) Remove blocking buffer from the wells  
j) Pipette 200 µl of standard curve solutions and samples (user-defined dilution with **REAGENT 4**) in duplicates in each well, and add 50 µl of CB 300 antibody test solution prepared in step h)  
k) Close the plate and incubate it for 16-24 hrs at 2-8°C (possible to do it over the week end)  
l) Empty the wells and wash 3x with washing buffer (**SOLUTION #3.4.**)  
m) Incubation with 200 µl of rabbit anti-mouse-HRP (DAKO P0260, 1:1000 in **REAGENT 4**) for 4-5 hrs at room temperature.  
n) Empty the wells and wash 3x with washing buffer (**SOLUTION #3.4.**)  
o) Wash 2-3x with dd. H<sub>2</sub>O.

#### **Enzyme reaction**

- p) Mix 1 volume of TMB-H<sub>2</sub>O<sub>2</sub> solution (**REAGENT 5**) with 20 volumes of substrate buffer (**REAGENT 6**)

21 ml of this solution is required per micro-well plate

This solution should not be vortexed (could cause precipitation) and has to be prepared just before use (stability 1 hr at 15-25°C)

- q) Pipette 200 µl of this TMB-H<sub>2</sub>O<sub>2</sub> solution per well  
r) Incubate 10 minutes at 15-25°C  
s) Stop the reaction by adding 100 µl stop buffer (**SOLUTION #3.3.**)  
t) Within 1 hr, analyze the plate with the photometer